Research Article

Mass Spectrometry Analysis Reveals Dynamic Changes of Protein Components in Lipid Rafts from Rat Liver after Partial Hepatectomy

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1. Abstract

 Lipid raft, as scaffolding platform for signal transduction, plays important role in the liver regeneration. But the lipid raft protein expression pattern during liver regeneration has been not reported. In this study, lipid raft proteins from the liver of 72 h post partial hepatectomy and sham-operated group were identified by liquid chromatography-tandem mass spectrometry (LC-MS/MS) combined with label-free semi-quantitative analysis. Totally 458 lipid raft proteins were identified, and most of identified lipid raft proteins were mainly involved in transporter, signal transduction, and metabolism. Moreover, label-free quantification analysis suggested that the level of 46 plasma membrane-related proteins have changed obviously (with ratio≥2) after 72 h hepatectomy. Several differently expressed proteins, including caveolin-1 and flotillin-1, were validated by western blotting. Further immuno fluorescence and quantitative real-time polymerase chain reaction data indicated the enhanced signal of flotillin-1 and caveolin-1 around the newly formed blood vessel. Here, the expression pattern of rat lipid raft protein after partial hepatectomy 72 h was investigated, and the differential expressed protein induced by partial hepatectomy was also indicated. Our results suggested that the increased expression of flotillin-1 and caveolin-1 could be involved in the liver regeneration.

2. Keywords: Liver regeneration; Lipid raft; Mass spectrometry; Proteomics; Flotillin-1

3. Introduction

Liver is unique in its ability to regenerate even in the mature stage [1, 2]. Liver regeneration is of great clinical significance in various liver-associated diseases [1]. It has been reported that liver regeneration undergoes three stages, including priming stage, proliferation stage and termination stage [3]. Angiogenesis is a key process in the third phase of liver regeneration. The key factor, Vascular Endothelial Growth Factor (VEGF), reached the peak of expression in 72 hours after Partial Hepatectomy (PH), which implicated that 72 h is [critical](file:///C:/Users/Canopus/Desktop/javascript:void(0);) node of angiogenesis during liver regeneration [4]. Besides transforming growth factor beta1 (TGF β1), as one of the regeneration terminators, remains elevated till 72 h to maintain appropriate size of liver [5]. Thus we focused on the time point of 72 h post Partial Hepatectomy (PH).

Lipid raft as thickened sections of the cell membrane plays an important role in cell signaling and cell-cell interaction [6, 7]. Caveolae, as a specified structure in the lipid raft, perform transport and signaling functions influencing cell growth, angiogenesis and transvascular exchange [8, 9].

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Caveolin-rich lipid rafts are involved in cholesterol trafficking, molecular transport and cell signaling [10-12]. It has been identified in the membranes of all liver cells, including parenchymal and non-parenchymal cells. More importantly, lipid rafts are active participants in multiple physiological and pathological conditions in individual types of liver cells [13]. Currently at least 300 proteins of lipid rafts in normal liver from human or rodents have been identified [14-17]. However, the protein functions of lipid raft in liver regeneration are not so clear. It is interesting to reveal the proteomic profile of lipid raft during liver regeneration after PH.

In this study, geLC-mass spectrometry (MS) /MS-based proteomic techniques have been applied to explore the protein composition of lipid rafts in rat liver. Furthermore, semi-quantitative analysis indicated the difference of proteins between the PH 72 h group and the sham-operated group (PH 0 h group). Among them, five proteins were verified by western blot and the distribution of flotillin-1 and caveolin-1 were proved by immunofluorescence.

4. Materials and Methods

4.1. Ethics Statement

This study was carried out in strict accordance with the guidelines established by the Committee on the Use and Care of Animals at the Hunan Province, P. R. China. The protocol was approved by the Ethics Committee of the University of Hunan normal university (Permit number: 010008). All efforts were made to minimize suffering. All the experiments were approved by and were conducted in the Committee.

4.2. Materials

Electrophoresis grade chemicals and Hyper film ECL reagent were obtained from GE Healthcare (Uppsala, Sweden). Colloidal Coomassie Brilliant Blue G-250, dithiothreitol and iodoacetamide were from Sigma Aldrich (St Louis, MO, USA). RC-DCTM kit was from Bio-Rad Laboratories (Hercules, CA, USA). Complete protease inhibitors cocktail was from Roche (Mannheim, Germany). Horseradish peroxidase conjugated anti-mouse IgG were obtained from BD Bioscience (San Jose, CA, USA). Anti-annexin A2, anti-Na+/K+ ATPase monoclonal antibody, anti-CD73 monoclonal antibody, anti-flotillin 1, anti-caveolin-1 were from Abcam (Cambridge, UK). Proteomics sequencing grade trypsin, and sucrose were obtained from Promega (Madison, WI, USA). Male Sprague-Dawley rats (averaging 200-250 g and 6-8 weeks old) were purchased from the Experimental Animal House at Centre South University (Changsha, China).

4.3. 2/3 Partial Hepatectomy (2/3 PH) Model

2/3 PH model was performed mainly as described by Higgins and

Anderson [2]. Briefly, rats were allowed provided with rat chow and water ad libitum and maintained under a 12/12 hours light/dark cycle before surgery. Ten rats were divided into PH 72 h group and a sham-operated group randomly so that each group included five rats. All the surgeries were performed between 8 am to 11 am. The median and left lateral lobes of livers from the PH group were ligated and resected. The sham-operated group was also subjected to midline incision with liver manipulation but without removal. All rats were maintained in a temperature-controlled room during the surgeries.

4.4. Isolation of Detergent-Resistant Lipid Raft Fraction from Rat Liver

Rat livers were dissected out from PH 72 h group and control group. After removed the gall bladder and blood vessels, the liver pieces $(3 g)$ were homogenized in eight times their weight $(m1/g)$ of cold solution A containing 50 mM HEPES, 1 mM CaCl₂ and protease inhibitors with a tissue-tearor (Biospec products, CE 2000, Mexico) until completely liquefied. After homogenization, samples were centrifuged at $600 \times g$ for 10 min at 4 °C firstly and supernatant were centrifuged at $100,000 \times g$ for 30 min at 4 °C. Then sediments from sham-operated and 72 h group were resuspended in 1 mL buffered saline (25 mM MES, 150 mM NaCl, pH 6.5, 1% Triton X-100) and mixed with equal volume of buffered 80% (w/v) sucrose. The membranes were transferred to a SW41 centrifuge tube and overlaid with 7.5 ml of buffered 30% (w/v) sucrose and about 2.5 ml of buffered 5% (w/v) sucrose. The discontinuous gradient was centrifuged at $200,000 \times g$ for 20 h in a SW41 rotor at 4°C. Twelve fractions were taken, starting from the bottom of the tube and the pellet at the bottom of the tube ("P" as the 13th fraction). All fractions were washed with buffered saline and centrifuged at $100,000 \times g$ for 90 min at 4 °C, with the pellet being resuspended in buffered saline [18].

4.5. Western Blotting

Protein fractions from rat liver were dissolved with $2 \times$ SDS sample loading buffer and total 50 μg separated on 10% separation gel and 5% stacking gel in each lane. Proteins were then transferred to nitrocellulose membrane and incubated with antibody at concentrations recommended by manufacturers. Images were taken by and Molecular Imager Gel DocTM XRS⁺ System and analyzed using Quantity One 1D-Analysis software (Bio-Rad, Hercules, USA).

4.6. In-Gel Digestion and Protein Identification

The entire lane was cut into 16 strips according to the visible bands. Each fragment was cut into 1-2 mm³ gel pieces. In-gel digestion was performed exactly as described before [19]. The trypsinized mixtures were analyzed on a Q-TOF microhybrid Mass Spectrometer (MicroTOFQ-II, Bruker Daltonics, USA) equipped with an ESI nanospray source. The digested peptides were injected on an Ultimate 3000 LC system (Dionex Ultrimate3000) and first desalted and pre-concentrated on a C18 PepMapTM pre-column (2 cm, ID 100 μ m, 5 μ m, C18 LC Packings). Peptides were separated on a C18 column (15 cm, ID 75 µm, 3 µm, C18) with a linear gradient of 5-40% solvent B (99.9 % acetonitrile with 0.1% formic acid) over 60 minutes with a constant flow of 300 nL/min. The peptides were detected in the positive ion MS mode and the data-dependent MS/MS mode. The data-dependent mode was used for survey scans (m/z 300 to 1500) in order to choose up to three most intense precursor ions. For collision-induced dissociation (CID) in MS/MS analysis, collision energies were chosen automatically as a function of m/z and charge. The collision gas was argon. The temperature of the heated sample source was 150 °C and the electro spray voltage was 1200 V. There were three technical replicates for each LC-MS run. The MS/MS data were acquired with the software compass control 1.3 (Bruker Daltonics, USA) that identified compounds and de-convoluted the spectra (Bruker compass Data Analysis 4.0 software) to "mascot generic files" (mgf), which include the mass values, the intensity (at least 5 counts/seconds) and the charge of the precursor ions. Background subtraction was set at 10%. Smoothing was done three times with "smooth window" (channels) 2.0 in the Savitzky Golay mode. The mgf. Files were analyzed using an online version of the Mascot 2.0 program (Matrix Science Ltd., London). Search parameters allowed for carbamido methylation of cysteine, oxidation (variable modification), one missed trypsin cleavage, and 0.5 Da mass accuracy for MS and MS/MS. To ensure the accuracy of protein identification, we selected the candidate proteins with individual scores greater than threshold according to the Peptide Summary Report of Mascot Search Results at a 95% confidence level (p <0.05).

4.7. Label-Free Semi-Quantitative Analysis

Around 2 μg sample from sham-operated and PH 72 group to perform a semi-quantitative analysis, respectively. A semi-quantitative analysis was performed for lipid raft proteomic data, as described previously [20]. A normalization protocol was used to normalize the spectral counts so that the total counts for each group were similar. The average spectral counts for each protein were generated for sham-operated and PH 72 group. The total spectrum number of peptides for each protein was detected to estimate the relative abundance of each protein. We chose protein with peptides no fewer than 2 as the confidence protein, and obtain differences of proteins between PH 72 and sham-operated group according to the exponentially modified protein abundance index (emPAI). Only those proteins with ratio≥2 are considered to be differential expressed proteins.

4.8. Bioinformatics Analysis

The Mascot score and number of unique peptides used to identify a protein as well as the sequence of each peptide were retrieved from Mascot searching output files. The mappings of putative trans membrane domains were carried out using the trans membrane hidden Markov model (TMHMM V 2.0) algorithm. The sub cellular location and function of the identified proteins were elucidated by literatures and gene ontology (GO) cellular component and function terms, respectively. The networks and pathways of the differentially expressed proteins were generated through STRING (Version 10.5, https:// string-db.org).

4.9. Fluorescence Immunohistochemistry

Immunohistochemistry was performed as described before²¹. Liver tissue was cut into around 0.5 cm³ pieces and fixed in 4% paraformaldehyde. After fixation, the tissue block was embedded in paraffin and sectioned in the sagittal plane. The mounted sections were de-paraffinized in xylene and rehydrated in a graded series of ethanol solutions. Epitopes were retrieved by heating sections in 10 mM sodium citrate buffer (pH 6.0). Sections were incubated in primary antibodies (diluted in phosphate-buffered saline plus 5% goat serum) overnight and then treated with a species-specific Alexa-Fluor 488 dye conjugated secondary antibody (Molecular Probes, Invitrogen) diluted in phosphate-buffered saline plus 5% goat serum at room temperature for 2 h. The immune labeled sections were examined under a Leica DM6000B microscope.

4.10. Quantitative Real-Time Polymerase Chain Reaction

Total RNA was harvested from about 100 mg of rat liver using TRIzol (Invitrogen, USA). First-stand complementary DNA (cDNA) were synthesized from 0.2 µg of total RNA for quantitative real-time polymerase chain reaction (qRT-PCR) (Invitrogen, Carlsbad, CA, USA). Quantitative real-time polymerase chain reaction (qRT-PCR) analyses were performed using the Light-Cycler system (Roche, Indianapolis, IN). PCR was performed using 12.5 µl of 2 X Master Mix SYBR Green I (Takara, Japan), 0.5 µl of each primer, and 1 µL of sample or H_2O to a final volume of 25 μ L. The amplification conditions were as follows: 95 °C (5 minutes); 40 cycles at 95 °C (15 seconds) and 65 °C (35 seconds); and extension. Detection of SYBR Green fluorescence at the end of the extension reflects the amount of double-stranded DNA. The expression of the target genes (Flotillin-1 and Caveolin-1) was calculated based on the ratio of the gene of interest to the glyceraldehyde 3-phosphate dehydrogenase (GAPDH) housekeeping gene. The primer set sense and antisense sequences for the genes were as follows: GAPDH forward, 5'-TGG AGT CTA CTG GCG TCT T-3'; GAPDH reverse, 5'-TGT CAT ATT TCT

CGT GGT TCA-3'; Flotillin-1 forward, 5'-AAG TTC TCA GAG CAG GTT TTC AAG-3'; Flotillin-1 reverse, 5'-TTC CCC AAA GAG TGT AAG TAA TCC-3'; Caveolin-1 forward, 5'-CCC CAA GCA TCT CAA CGA C-3'; Caveolin-1 reverse, 5'-GGT AGA CAG CAA GCG GTA AAA C-3'.

5. Results

5.1. Subcellular Fractionation and Identification of Lipid Rafts

An enriched lipid raft fraction was isolated from plasma membrane preparations using Triton X-100 solubilization and discontinuous

sucrose density gradient centrifugation (Figure 1A). This approach allowed the insoluble lipid rafts (detergent-resistant micro domains, DRMs) to float to the 5%/30% sucrose boundary, where they were observed as a light-scattering band (Figure 1B). Proteins of plasma membrane and soluble fractions from rat liver were compared with DRMs proteins using immuno blotting. As shown in (Figure 1C), lipid raft marker proteins flotillin-1 and caveolin-1 [22, 23] have been enriched within the DRM fraction. This clearly demonstrated the enrichment of lipid raft fraction relative to plasma membrane was enriched by such methods.

Figure 1: Sucrose density gradient enrichment of detergent-resistant membranes from rat liver. (A) Diagrammatic representation of sucrose density gradient fractionation of plasma membranes. (B) Photograph of tube showing the presence of a light-scattering band of DRMs at the interface of the 5% and 30% sucrose layers, corresponding to fractions 3 and 4. (C) Western blots of isolated protein fractions from rat liver. Blot developed with Caveolin-1 antibody and flotillin-1 antibody.

DRM proteins from sham-operated group and PH 72 h group were separated on SDS-PAGE. As shown in (Figure 2), compared with PH 0 h group, there were 7 obviously different bands in PH 72 h group. Sixteen strips on the gel were cut in parallel and subjected into enzyme digestion and mass spectrometry.

Figure 2: Representative SDS-PAGE profile of lipid rafts proteins in rat liver after 2/3 hepatectomy. LRs proteins from sham-operated and PH 72 groups were separated on SDS-PAGE. Intensity of bands indicated by pentagram exhibited obvious differences between PH72 group and sham group after partial hepatectomy.16 strips on the gel were cut in parallel and subjected into enzyme digestion and mass spectrometry.

5.2. Proteomic Analysis of Lipid Rafts From Rat Liver; Were Present in All the Secretome Preparations

In total, 458 proteins were identified in lipid raft from PH 72 h group and control group (Supplementary (Table 1), and totally 316 proteins were present in both groups (Figure 3A). The identified proteins were characterized on the basis of their sub cellular localization, as predicted from the Gene Ontology (GO) terms or descriptions provided in Rattuse entries. Among the 456 identified proteins, 430 had descriptions for their sub cellular location. The identified proteins were associated with plasma membrane (27.5%), mitochondria

 (26.6%) , cytoplasm (11.6%) , and ribosome (11.4%) . Besides, there were proteins located in endoplasmic reticulum (11.1%), nucleus (3.1%), and peroxisome (1.5%). And still a relatively small proportion (7%) was unclassified (Figure 3B). We also classified the 458 proteins according to their GO function description. 28.2% were annotated as ion channel and transporters, 22.7% as catalytic proteins, 19.9% as structural proteins, 11.8% as regulatory proteins, 4.8% as signal proteins and receptors, 2.4 as cell adhesion proteins. Other annotated proteins (10.9%) included some unknown proteins (Figure 3C).

Figure 3: Classification of the total lipid raft proteins identified in rat liver after partial hepatectomy. (A) A Venn diagram of based on the comparison of lipid raft proteins from PH 72 h group and control group. (B) Classification of the identified lipid rafts proteins based on subcellular localization information. (C) Functional classification of the identified lipid rafts proteins.

Furthermore, the molecular weight, isoelectric point, hydrophobicity value and trans membrane region were analyzed for all proteins (Figure 4). Most of them were less than 60 kDa but there was nearly 7% molecular weight higher than 100 kDa (Figure 4A). The isoelectric points of these proteins ranged from 4 to 10 and falled into normal distribution pattern (Figure 4B). The proportion of "neutral proteins" ($6 \le pI \le 8$) is abundant. Meanwhile, 277 trans membrane proteins were found and 30 proteins have at least 10 trans membrane domains (Figure 4C) and the hydrophobic value of near 3.2% protein is more than 0.5 (Figure 4D).

Figure 4: Physicochemical characterization of proteins identified in lipid rafts of rat liver. Molecular mass (A), isoelectric point (B), number of transmembrane domains (TMD) (C), and GRAVY value (D).

5.3. Differentially Expressed Lipid Raft Proteins

In total, 377 and 447 proteins were identified in the sham-operated and group PH 72 group, respectively. And 46 plasma membrane-associated proteins were found to be differentially expressed (with change more than 2 folds) (Supplementary (Table 2). Compared with the sham operation group, 34 proteins were up-regulated whereas 12 proteins were down-regulated at PH 72 h group. All the 46 differentially-expressed proteins were pooled together for pathway analysis using STRING. Among them, up to 36 proteins could be clustered into cav-1 protein-mediated pathway (Figure 5). Obviously, cav-1 protein was one of the important nodes at all the networks and modulated the diverse physiological processes, such as material transport, signal transduction and [cytoskeleton](file:///D:\Youdao\Dict\7.0.1.0214\resultui\dict\result.html?keyword=cytoskeleton) and so on. Also as a mark protein of lipid raft, flot-1 has a close interaction with cav-1. According to result of pathway analysis, besides cav-1 protein, we speculate that the flot-1 also is an important factor involved in the events at PH 72 h.

Table 2: Changes of Protein Expression

Figure 5: Construction of integrated signaling networks. Data sets obtained from proteomic analysis were uploaded to the web-based tool STRING to identify functional signaling networks. The protein-protein interaction network is presented by Gene Ontology classification. The corresponding proteins for the genes are indicated in supplemental tables 2.

5.4. Validation of Differential Protein Expression

Western blot was conducted to validate the differential expression and five differential proteins were included in present study, such as Nt5e, ATP1A-1, annexin A2, caveolin-1 and flotillin-1. As shown in (Figure 6), the expression level of Nt5e, Annexin A2, caveolin-1 and flotillin-1 in lipid raft increased in the PH 72 h when compared with that of control (*P<*0.01 for Nt5e and Caveolin-1; *P<*0.05 for Annexin A2 and Flotillin-1). In contrast, the abundance of ATP1A-1 in lipid raft after PH 72 h decreased (*P<*0.05 *vs.* PH 0 group) from western blotting. Western blot results of caveolin-1 and flotillin-1 were in accordance with the data from label free quantitative assay. But results of Nt5e, ATP1A-1, annexin A2 is different to label free quantitative assay which might link to that label free quantitative assay is semi-quantitative research. In addition, reproducibility of proteins might also be affected by physiological factors. Therefore, there will be difference between western blotting and quantitative results.

To further validate of these differential proteins expression, the messenger RNA (mRNA) levels of those molecules were determined by qRT-PCR at several time points (Figure 7A-E). These results were in accordance with the data from western blot assay. Besides, the mRNA level of some plasma membrane proteins, which expressed more than 5 times in PH72 /SHAM group, was determined by qRT-PCR, such as flotillin-2, abcb4 and xpnpep2. These results were in accordance with the data from label free quantitative assay (Figure 7F-H).

Figure 6: Western blotting and qRT-PCR analysis for Caveolin-1 and Flotillin-1. (A, B) The Western blotting results for Nt5e, Atp1a1, Annexin A2, Caveolin-1 and Flotillin-1. Sham-operation control groups and after PH groups. Each panel is the representative result of three independent experiments.

Figure 7: qRT-PCR analysis. (A-H) Nt5e, Atp1a1, Annexin A2, Caveolin-1, Flotillin-1, Flotillin-2, Abcb4 and XPnpep2 gene expression in rat liver of PH 24 h, 48 h, 72 h and sham groups by qRT-PCR.

5.5. Illustration of Immunofluorescence and Qrt-PCR Analysis of Caveolin-1 and Flotillin-1.

We speculated that differentially expressed lipid raft protein flotillin-1 and caveolin-1 play important roles in liver regeneration. Then immunofluorescence was preformed to observe the localization of flotillin-1 and caveolin-1 during liver regeneration. As shown in (Figure 8), the signal of flotillin-1 and caveolin-1 is very weak at sham-operated group whereas the scattered bright signals emerged in PH 24 h, especially around the small blood vessel liked structure. From PH 48 h, the signals become stronger and widely distributed and PH 72 h have brightest field under microscope. All above data suggested the increased expression of flotillin-1 and caveolin-1 with the progression

of live recovery after PH (Figure 8). Besides, we can also find caveolin-1 and flotillin-1 expressed on endothelial cells of new vessels at PH 24 h and PH 36 h and then they mainly expressed on hepatocyte.

To further study the effect of caveolin-1 and flotillin-1 on liver regeneration, the messenger RNA (mRNA) levels of those molecules were determined by qRT-PCR at several time points (Figure 7D, E). Compared with sham operation group, the mRNA expression level of flotillin-1 was gradually raising and caveolin-1 also had significantly higher levels after a short down-regulation at PH 24 h, which showed a consistent trend with caveolin-1 and flotillin-1 on protein level. These results indicated that caveolin-1 and flotillin-1 might play important roles during liver regeneration.

Figure 8: Expression of flotillin-1 and caveolin-1 in liver after after PH. The signals of flotillin-1 and caveolin-1 in sham-operated group (PH 0) and after PH groups (PH 24, PH 36, PH 48, PH 72) were illustrated by immunofluoresence. The nucleus were stained with Hoechst as shown in blue. Fluorescence was observed using a Nikon EclipseE600 microscope. The right two columns of immune fluorescence spectrum is local enlarged drawing of the corresponding white box in second and third column. The arrows "1" indicate hepatic sinusoid within green fluorescence, and arrows "0" indicate hepatic sinusoid without green fluorescence Scale bars, 100 μm.

6. Discussion

Recently, the progress of liver regeneration has been widely studied through proteomics and transcriptomics [24-27]. Among them, most of the studies focus on the change of molecules at early stage, i.e., 24 h after PH. In this period, liver cell proliferation is the major event and cell cycle-related proteins were identified frequently. However, only a few reports were focused on the late stage of liver recovery, i.e., 72 hours after PH [19, 28-29].

Our previous study has been revealed the proteomic profile of liver sinusoidal endothelial cells (LSECs) in rat liver [19]. Present study was concentrated on the variation of lipid raft proteins during liver regeneration. The lipid raft proteins were separated on 1D SDS-PAGE and identified by LC-MS/MS. According to available published data, this lipid raft proteome is the largest one in rat liver. It need to be mentioned that it still have contamination from cytosolic proteins (around 11.6%), nuclear proteins (3.1%) and ribosomal proteins (11.4%) in this work. Given the known sub cellular localization, those proteins did not affect analysis of lipid raft proteins. By using the same method, there were 26.52% cytosolic proteins included in lipid raft data [30]. And 28.72% cytosolic proteins in the lipid raft data [31]. It might indicated that the contamination from cytoslic proteins or other organells is difficult to remove by sucrose gradient ultracentrifugation and further modification is worth to be explored for pure preparation of lipid raft.

Our previous work on the profile of plasma membrane covered partial information of lipid rafts [19]. Among them, 98 proteins were identified repeatedly in this study. In addition, the profile was identified by including 196 proteins in total and 90 proteins were also detected in this study [14]. Proteins expression level between PH 0 h and PH 72 h were analyzed by label free quantification strategies, and 46 proteins changed dramatically during partial hepatectomy and were involved in liver regeneration in this study, especially angiogenesis. For molecules especially expressed at after PH 72 h, [28] identified 218 proteins in total, 83 of which presented over 2-fold differentially expressing level. Among them, 20 proteins were included in our profile of 72 hours after surgery. In addition, the transcriptomics data after PH 72 h identified 767 transcripts, which might be corresponding to 767 proteins and 30 proteins of them were also presented in our study. Among 30 transcripts, the corresponding proteins of one mRNAs, i.e., monocyte differentiation antigen CD14 showed similar pattern with this study [29].

As key molecules of lipid rafts, flotillin-1 and caveolin-1 play important roles in different physiological and pathological conditions. Caveolin-1 is essential molecular in cancer occurrence, development, invasive metastatic and prognosis [32, 33]. It has been reported that caveolin-1 is essential for liver regeneration, which mainly functions in the formation of the lipid droplets and new membrane, and also energy supply [34, 35]. The expression intensities of caveolin-1 was mainly localized at LSECs and hepatic stellate in liver cancers and para-cancer liver tissues [36]. In this work, the expression of caveolin-1 was upregulated slightly on plasma membrane of LSECs after PH, which confirmed the importance of caveolin-1 and also illustrate the reliability of our data.

Flotillin-1 could provide a platform for the interaction between proteins in lipid raft and involve in vesicular trafficking and signal transduction [23, 37-38]. Besides roles in the biomembrane, flotillin-1 might participate in development and progression of tumor genesis and angiogenesis [39-41].The study has confirmed flotillin-1 was over expressed in Hepatocellular Carcinoma (HCC) and indicated flotillin-1 played an essential role in HCC [42]. Furthermore, other report indicated that the hyper expression of caveolin-1 and

flotillin-1 could contribute to development of HCC though active Toll-like receptors signaling [43], but it was not clear whether parenchymal cells proliferation was dependent on flotillin-1 in liver. Here, our data suggested that flotillin-1 might play a role in liver regeneration. It could be also speculated from the interaction network that flotillin-1 may provide signal transduction platform for angiogenesis factor such as integrin β1, vascular endothelial growth factor-A and fibronectin-1. But it has not been reported that flotillin has relation with LSECs. And molecular mechanisms of flotillin-1 in the angiogenesis, especially liver regeneration, need to be further studied.

In addition, The trends of flotillon-1 expression was consistent with caveolin-1 in liver cancers and paracancer liver tissues [43] and had slightly up-regulated in this work. We speculate that the differential expression of caveolin-1 and flotillin-1 might be one of the differences between normal liver regeneration and liver cancer.

In summary, our data presented a large lipid raft proteome database. Furthermore, differential expression profile after PH 72 h and sham-operated group in liver were also revealed. Among them, flotillin-1 and caveolin-1 as scaffold proteins could play important roles in angiogenesis, which may provide further insight into the mechanism of angiogenesis and the regulation of liver regeneration.

7. Acknowledgment

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